SM08502, a novel, small-molecule CDC-like kinase (CLK) inhibitor, demonstrates activity against cancer stem cell (CSC)-enriched pancreatic cancer cells and suppresses stemness in vitro

Carine Bossard¹, Nathalia Cruz¹, Brian Eastman¹, Chi-Ching Mak¹, Sunil KC¹, Betty Tam², Timothy Phalen¹, Steven Cha¹ ¹Samumed, LLC, San Diego, CA; ²Formerly Samumed, LLC, San Diego, CA

Background

- Cancer stem cells (CSCs) are a rare subpopulation of quiescent tumor cells with stemness, the ability to self-renew and form new tumors, and may contribute to chemotherapy resistance, proliferation, and relapse in pancreatic cancer (PC)^{1,2}
- Aberrant activation of the Wnt signaling pathway is implicated in multiple cancer hallmarks including proliferation, metastasis, and immune evasion, as well as the maintenance and survival of CSCs^{1,3}
- CDC-like kinases (CLKs) phosphorylate serine/arginine-rich splicing factors (SRSFs), which regulate spliceosome assembly and subsequent gene expression^{4,5}
- SM08502 is a novel, oral, small-molecule pan-CLK inhibitor that has been shown to potently inhibit Wnt pathway activity in preclinical colorectal cancer models⁶
- These studies examined the ability of SM08502 to impair CSC viability and stemness in PC cell lines

Methods

• Panc-1 cell cultures were enriched in CSCs (Panc1-CSC) by inducing anoikis, programmed cell death triggered by non-adherent growth conditions (Fig. 1):



- •Cell (spheroid) viability Panc1-CSC cultures were plated on day zero (D0) in ULA plates and incubated until D3. On D3, the nascent spheroids were treated with salinomycin, napabucasin (both CSC-inhibiting positive controls^{7,8}), or SM08502 until D6, and viability was assessed via CellTiter-Glo 3D[™] luminescence assay (**Fig. 2**)
- •Spheroid formation Panc1-CSC cultures were plated on day zero (D0) in ULA plates and treated on D1 with salinomycin, napabucasin, or SM08502 for 6 days, after which the plates were imaged using a CellInsight[™] CX5 imager (**Fig. 3**)
- Gene expression qRT-PCR after 20 h exposure to vehicle or SM08502 (1 μ M) using TaqMan[®] primers and normalizing expression to GAPDH (via $^{\Delta\Delta}Ct$); stemness-associated genes were assessed in both Panc-1 and Panc1-CSC cultures (**Fig. 4**)
- Spheroid-forming frequency HPAFII, Capan-1, and Panc-1 parent cell cultures were plated on D0 in 6-well plates and treated with SM08502 (1 µM) or vehicle per the timeline below. Spheroid-containing wells were counted around D24 (**Fig. 5**)

Pla	te parental cells		+ SM08502 or vehicle				Fresh med	ia without S		Plate at 1 or 10 cells/well	
	1	I	I	l	l	l	I	I	I		in ULA 96w plate
Day	0	1	2	3	4	5	6	7	8	9	Count spheroids around D24



Figure 2. SM08502 potently impaired Panc1-CSC spheroid viability



	SM08502	Salinomycin	Napabucasin
EC ₅₀ (μΜ)	0.1028	0.2682	2.301
Span	1.031	0.8255	0.9032

Figure 3. SM08502 impaired formation of Panc1-CSC spheroids

	DMSO		0.0005	0.0015	0.0046	0.0137	0.0412	0.1235	0.3704	1.111	3.333	10	u.
SM08502			•	Me		ð			Ŷ			D. M	
	The .					-			ta		***		
	ġ.	×								-	4		
	Å.		- Maria	÷.			4	**	A				
Napabucasin		in the second			a la	ýø				0	166		
	1						-				A		
			· 🍂		•				38			*	
	- M		ð		-				ø			and a second	
Salinomycin				· ()		As	i de			*	all a		
			-		¢				A	- AN			
	-	1				R			4				
	Ņ						4		4	· ·			

Results



*p<0.05 vs. Panc-1 DMSO [†]p<0.05 vs. Panc1-CSC DMSO





Poster #C08

Conclusions

- **CSCs were successfully enriched in Panc-1 parent** cell cultures
- SM08502 demonstrated strong activity against CSCs in pancreatic cancer cell lines
- SM08502 anti-CSC activity was more potent than other CSC inhibitors (salinomycin and napabucasin) in vitro
- SM08502 inhibited the stemness of CSCs and parental PC cells
- SM08502 can potentially address relapse and treatment resistance in PC by depleting CSCs and reducing stemness in tumors
- A Phase 1 study assessing the safety, tolerability, and pharmacokinetics of SM08502 in subjects with advanced solid tumors is ongoing (NCT03355066)

References

- . Visvader J and Lindeman G. Cell Stem Cell. 2012;10:717-28.
- 2. Li C, et al. Cancer Res. 2007;67:1030-7.
- 3. Ducharte Y, et al. Crit Rev Oncol Hematol. 2016;99:141-49.
- 4. Colwill K, et al. EMBO J. 1996;15:265-75.
- 5. Long JC, et al. Biochem J. 2009;417:15-27
- 6. Bossard C, et al. J Clin Oncol. 2019(supp;abstr e15185).
- 7. Naujokat C and Steinhart R. Journal of Biomedicine and Biotechnology. 2012;950658.
- 8. Li Y, et al. *PNAS*. 2015;112:1839-1844.

All authors are employees, shareholders, or consultants of Samumed, LLC. Other disclosures are listed in the published abstract.

